

Published in final edited form as:

*Clin Immunol.* 2013 December ; 149(3): . doi:10.1016/j.clim.2013.02.003.

## Anti-Cytokine Therapies in T1D: Concepts and Strategies

Gerald T. Nepom, MD, PhD<sup>a</sup>, Mario Ehlers, MD, PhD<sup>b</sup>, and Thomas Mandrup-Poulsen, MD<sup>c</sup>

Gerald T. Nepom: nepom@benaroyaresearch.org; Mario Ehlers: mehlers@immunetolerance.org; Thomas Mandrup-Poulsen: tmpo@sund.ku.dk

<sup>a</sup>Benaroya Research Institute, 1201 Ninth Av, Seattle, WA 98101 USA

<sup>b</sup>Immune Tolerance Network, 185 Berry St, #3515, San Francisco, CA 94107 USA

<sup>c</sup>University of Copenhagen, Blegdamsvej 3, 2200 Copenhagen N, DENMARK

### Abstract

Therapeutic targeting of proinflammatory cytokines is clinically beneficial in several autoimmune disorders. Several of these cytokines are directly implicated in the pathogenesis of type 1 diabetes, suggesting opportunities for design of clinical trials in type 1 diabetes that incorporate selective cytokine blockade as a component of preventative or interventional immunotherapy. The rationale and status of inhibitory therapy directed against IL-1, TNF, IL-12, IL-23, and IL-6 are discussed, towards a goal of using cytokine inhibition as a therapeutic platform to establish an in vivo milieu suitable for modulating the immune response in T1D.

### Keywords

Immunotherapy; autoimmunity; regulatory T cells; anti-inflammatory

### 1.0 Overview

Clinical trials with targeted immunotherapy directed towards lymphocytes in intervention trials, using anti-CD3 (teplizumab or oteplizumab), anti-CD20 (rituximab), or CTLA4-Ig (abatacept), resulted in transient maintenance of insulin-secreting function in some type 1 diabetes (T1D) subjects. However, the overall picture is not a positive one, because the majority of treated subjects reverted to a pattern of progressive beta cell loss after treatment [18;22;35;38], indicating the recurrence of pathogenic autoreactivity. Recurrence of pathogenic lymphocytes has also been seen in studies of T1D subjects who receive pancreas transplants, in which autoreactive effector lymphocytes have been shown to survive in spite of aggressive immunosuppressive therapy, and in some cases expand in tandem with recurrence of T1D in the transplanted organ graft [54]. These accumulated clinical observations need to be addressed in the next generation intervention trials in T1D, as they indicate a compelling need to address not only the specific effector populations, but also the host environment that nurtures and directs the recurrent immune response after initial therapy.

© 2012 Elsevier Inc. All rights reserved.

Corresponding author: Gerald T. Nepom, MD, PhD, Benaroya Research Institute, 1201 Ninth Avenue, Seattle, WA 98101-2795 USA, Tel: 206-342-6515, Fax: 206-342-6581, nepom@benaroyaresearch.org.

**Publisher's Disclaimer:** This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Potential strategies for achieving a more durable immunological effect can be categorized as:

- i. Enhancing lymphodepletion or immunomodulation, i.e., making some of the current therapies more effective through synergistic mechanisms of action;
- ii. Boosting or replacing dominant regulatory elements in the immune system that will create a sustainable balanced immunological profile after effector cell depletion or modulation therapy;
- iii. Changing the tissue microenvironment to alter the likelihood that recurrent effector pathways will be activated;
- iv. Creating a cytoprotective milieu supporting beta cell survival and repair.

In this brief review, we will discuss an approach towards achieving these strategies through manipulation of specific cytokine pathways, with examples focused on blockade of IL-1, TNF, IL-12, IL-23, and IL-6.

## 2.0 Cytokine Therapies

### 2.1 Anti-interleukin 1 (anti-IL-1)

The family of interleukin-1 (IL-1) proteins consists of five main groups of agonists, partial agonists, and antagonists, some of which have arisen from gene duplication within the IL-1 gene cluster on chromosome 2 [10] (Table 1). These molecules are evolutionarily highly conserved and constitute key mediators of innate immunity in primitive organisms, dating back to starfish. The subgroup-agonists activate distinct receptors but with wide overlap in the use of the IL-1 receptor accessory protein (IL-1RAcP) as co-receptor, opening up the potential for the therapeutic targeting of IL-1, IL-36, and IL-33 action by the same anti-IL-1RAcP biologic.

In addition to the complex and branched control on IL-1 signalling provided by the existence of receptor antagonists and decoy receptors, the action of the IL-1 family of proteins is subjected to pronounced transcriptional, translational, and posttranslational regulation. All nucleated cells investigated so far have been found capable of expressing IL-1 family proteins upon appropriate stimulation, including the pancreatic beta cell. Monocyte-derived and dendritic antigen-presenting cells (APCs) are the most potent IL-1 producers in response to a wide variety of stimuli. Of note, the production of mature IL-1 depends upon a two-signal sequence: signal I is induced by several activators of the canonical NF  $\kappa$ B signalling pathway, such as TLR ligands, metabolic factors, and cytokines that engage receptors recruiting the intracellular MyD88 docking protein. Signal I is required to drive proIL-1 mRNA transcription and translation, and this signal is amplified by glucose-induced calcium-, ERK MAP kinase-, and ROS-dependent pathways. However, proIL-1 is biologically inert and needs to be processed by caspase-1 cleavage.

Inactive pro-caspase 1 is activated by cleavage induced by signal II, which is conferred via a multiprotein complex named the inflammasome, a group of intracellular receptors of danger-associated molecular patterns (DAMPs) [30]. How the inflammasome is activated and which ligands bind to the ligand-sensing, leucine-rich domain is incompletely understood, but diverse extracellular stimuli, such as ATP, nutrients and metabolic factors, and non-degradable particulates (cholesterol or uric acid crystals, amyloid), which elicit a process of frustrated phagocytosis seem to converge on the generation of reactive oxygen species that lead to dissociation of the thioredoxin inhibitory protein TXNIP from thioredoxin. TXNIP has been proposed to activate the inflammasome [36;58]; alternatively ATP-stimulated potassium efflux via purinergic receptors may be sensed by the

inflammasome as activating signal. The expression of the inflammasome components is also influenced by signal I.

In contrast to the detailed insights into the regulation of IL-1 expression and processing, little is known about how IL-1, which lacks a leader sequence for secretion, is exported out of IL-1-producing cells and how this process is regulated. However, the better understood, intricate regulation of IL-1 offers multiple possible targets for intervention.

As most cells synthesise IL-1, virtually all cells studied hitherto express IL-1R and respond to IL-1. The main action of IL-1 is to drive the acute phase response of inflammation and stress, but IL-1 has multiple, additional neuronal, endocrine, metabolic, and immune effects, including effector T-cell co-stimulation and inhibition of regulatory T-cell function [11]. IL-1 operates at the top of the cytokine and chemokine hierarchy and drives the expression of multiple proinflammatory and anti-inflammatory cytokines and chemokines, including the expression of IL-1 itself, and is in turn regulated by multiple other cytokines. IL-1 signals mainly via the NF- $\kappa$ B and MAPK pathways but also via small G proteins and other pathways only partially understood. The cellular effects involve changes in gene expression and protein activity to assist cell and host defense, tissue repair, and remodelling, as well as cellular stress and destruction via endoplasmic reticulum and mitochondrial stress pathways.

The many ligands and receptors of the IL-1 family offer a wide portfolio of opportunities for intervention [9]. Apart from recombinant IL-1RA, soluble IL-1TI or II receptor and an IL-1TI R-IL-1RAcP fusion protein (the so-called IL-1 trap), several antibody-based antagonists are manufactured and marketed, including anti-IL-1, anti-IL-1TI R, and IL-1RAcP antibodies.

Apart from its immunoregulatory properties, IL-1 has long been known to exert profound inhibitory, cytostatic, pro-necrotic, and pro-apoptotic effects on the pancreatic beta cell [29]. IL-1 is expressed early in the insulinitis infiltrate and may be a circulating biomarker of T1D risk. However, whereas anti-IL-1 antagonism has shown efficacy in preclinical models of T2D and reduces glycemia via improved beta cell function in T2D patients [7;12;25], anti-IL-1 strategies or genetic ablation of IL-1 or receptor have shown modest or no protective efficacy in animal models of T1D [29]. In contrast, IL-1 antagonists strongly synergize with suboptimal anti-CD3 monoclonal antibody (mAb) therapy to accelerate and promote reversal of overt diabetes in the nonobese diabetic (NOD) mouse [1].

Only one small unblinded non-randomised study of IL-1 antagonism with IL-1RA (anakinra) in 15 recent-onset T1D children has been published [47], showing reduced insulin requirements and insulin-adjusted glycosylated haemoglobin compared with two historical control groups, an effect that was not confirmed in two yet unpublished randomized placebo-controlled trials.

Thus, although there is solid preclinical rationale for IL-1 as an interventional target in T1D, results have been disappointing so far. It is possible that timing and dosing of IL-1 antagonists are critical parameters as is the use in combination with other anti-cytokine or anti-adaptive or innate immune cell approaches [1].

## 2.2 Anti-tumor necrosis factor (anti-TNF)

There are nineteen known members of the TNF family, which bind to specific receptors with limited cross-binding [48] (Table 2). The prototypic member of this family is TNF. In contrast to IL-1, proTNF contains a leader sequence, but the proTNF is inserted into cell and plasma membranes as a homo-trimeric complex (membrane-bound TNF), which can be shed by the action of a membrane metalloprotease, TNF-converting enzyme (TACE), or

ADAM17. Shedding probably occurs both from membranes in the *trans*-Golgi and from the plasma membrane. How shedding is regulated is incompletely understood, but it is known to be subject to inhibition by tissue inhibitor of metalloproteinase 3 (TIMP-3).

TNF is produced by many cells, including macrophages, NK cells, CD4 T cells, endothelial cells, and adipocytes, in response to LPS and other bacterial products, and IL-1 and other cytokines. TNF binds to the homo-trimeric TNF receptor, which recruits intracellular adaptor proteins, such as TRADD, FADD, and TRAF2. Via the death domains of FADD, caspase 8 is activated which in turn triggers the effector caspase of apoptosis, caspase 3. TRAF2 elicits NF- $\kappa$ B and MAPK activation, redundant to IL-1 signalling.

TNF was discovered as a tumor apoptosis- and cachexia-inducing cytokine, and, like IL-1, is a potent mediator of the acute phase response and septic shock. In addition, like IL-1, TNF is a chemokine and an adipocytokine secreted from adipose tissue and involved in the pathogenesis of insulin resistance and the metabolic syndrome. Although TNF alone has little direct effect on the pancreatic beta cell, it synergizes potently with IL-1 and IFN- $\gamma$  in inducing beta cell dysfunction and apoptosis [28].

The effects of TNF or TNF antagonism in T1D animal models have been conflicting. Although expression of TNF locally in the pancreatic islets under the rat insulin promoter accelerates T1D by inducing a florid islet inflammatory reaction, TNF or TNF blockade may both protect and aggravate diabetes development depending upon dose and timing [27].

Only one clinical study of blocking TNF in recent-onset T1D has been conducted. This was a 24-week, double-blind, placebo-controlled phase 2a clinical trial [32]. Out of almost 400 eligible patients aged 3–18 and identified in a 5-year study period, it was possible to randomize only 18 subjects with a mean age of 12.5 to the recombinant, soluble, neutralizing TNF receptor-IgG fusion protein etanercept; 17 completed the follow-up protocol, and the study was terminated due to slow recruitment. Etanercept-treated patients achieved a 0.2% lower glycated hemoglobin between 8 and 24 weeks of follow-up; this difference was statistically significant and persisted 12 weeks after withdrawal. C-peptide rose in 6/9 etanercept-treated versus 1/8 placebo-treated subjects, and the mean C-peptide was significantly higher in the etanercept arm. The improved beta cell function translated into a significantly lower insulin requirement in the active arm. There were no evident safety concerns in this small sample of subjects. Larger studies are needed to confirm these encouraging results. However, slow recruitment, despite a large proportion of eligible subjects (not randomized because information about the study resulted in lack of consent) questions the feasibility of using this approach.

### 2.3 Anti-IL-12/23

IL-12 and IL-23 are secreted by dendritic cells, macrophages, and monocytes and support the maturation and maintenance of proinflammatory effector T cells. Their primary roles appear to be involved in the induction of committed T cell lineages, IL-12 for Th1 and IL-23 for Th17 cells, acting on naïve, immature, and/or “plastic” uncommitted T cells in concert with stimuli that include antigen exposure and other cytokines. Because IL-12 and IL-23 are heterodimers that share the common p40 subunit, inhibition of p40 is an attractive target to interfere with both Th1 and Th17 development and function.

Ustekinumab is a human IgG1 monoclonal antibody that binds with high affinity and specificity to the shared p40 subunit in both IL-12 and IL-23, blocking signaling by inhibiting the interactions of these cytokines with their receptors [4;14]. Ustekinumab has shown efficacy in several human autoimmune diseases, including psoriasis, psoriatic arthritis, and Crohn’s disease [8;26;37;43;56] for the treatment of adult patients (18 years or

older) with moderate to severe plaque psoriasis. Analysis of tissue from the affected target in those diseases (e.g., skin in psoriasis) has documented the expected mechanism of action for this therapy, with significantly diminished Th1 and Th17 activity after systemic administration of drug [46].

Rationale for use of ustekinumab in T1D is largely based on the attractive mechanistic rationale of blockade of both Th1 and Th17 pathways, since evidence from analysis of lymphocyte phenotypes in patients supports potential roles for each. In addition, since interruption of IL-12/23 signaling cascades interferes with production of downstream inflammatory mediators, such as IFN $\gamma$  and IL-6, the milieu for supporting Treg activity is improved. Most of the evidence implicating the Th1 and Th17 pathways in T1D is necessarily indirect, derived from analysis of lymphocytes and cytokine profiles from peripheral blood. Islet antigen-specific T cell lines and clones are readily expanded in cultures of peripheral blood lymphocytes from T1D subjects and produce multiple cytokines, notably IFN $\gamma$ . More recent reports indicate that peripheral blood CD4 cells from new-onset T1D subjects also produce IL-17 in response to activation or autoantigen stimulation, and interestingly IL-17 enhances beta cell apoptosis induced by other inflammatory cytokines, such as IFN $\gamma$  and IL-1 [2;20]. This has led to the proposal that signaling by early inflammatory mediators secreted by Th1 cells and macrophages renders cells susceptible to IL-17-mediated apoptosis [2]. This view suggests a fairly comprehensive picture in which inflammatory signals from innate immune cells (macrophages and dendritic cells), an autoreactive cytolytic response driven by adaptive Teff cells (Th1 and Th17), and an inadequate or dysfunctional protective response from Treg cells combine into a pathogenic program dependent on the IL-12/23 signaling pathways.

Studies in murine models are problematic for prediction of therapeutic efficacy for ustekinumab. In the NOD mouse the dominant Teff cells appear to be IFN $\gamma$ -producing Th1 cells. The role of Th17 cells in the NOD model is controversial. While some studies have shown a role for IL-17 and IL-21 in the development of T1D in the NOD mouse [15;49], others have suggested that the role of Th17 is indirect, possibly by conversion to Th1 cells [3;21]. The role of IL-12 blockade is similarly controversial in the NOD mouse—and contradictory. There have been suggestions that therapeutic efficacy may be dependent on the age of the mice, the stage of disease, and the type of disease model [17;34;40;41;51–53;55;57].

## 2.4 Anti-IL-6

IL-6 is an abundant proinflammatory cytokine associated with immunity and autoimmunity, produced by many different cell types. It is often characterized as “downstream” of IL-1, but in fact is involved in multiple, different stages of immune response and is a uniquely attractive therapeutic target. IL-6 plays an important role in the communication between the innate and adaptive immune systems, functioning to influence the development and action of both pathogenic and regulatory T cells. One particularly intriguing role for IL-6 is a pivotal position in Th17 and Treg commitment, in which IL-6 combines with TGF $\beta$  to promote the Th17 lineage, whereas, in the absence of IL-6, TGF $\beta$  contributes to Treg development [5;23]. In addition, IL-6 is a potent effector cytokine, with multiple direct tissue effects that promote proinflammatory cascades [33;39]. Thus, IL-6 blockade may promote both anti-inflammatory and pro-regulatory mechanisms simultaneously in a tissue environment under active autoimmune attack.

Studies in murine autoimmunity suggest that Treg function is disabled by inflammatory cytokines in the local microenvironment, which drive Teff lineages and inhibit Foxp3. For example, IL-6 induces methylation of the *Foxp3* gene enhancer in nTreg cells, leading to down-regulation of Foxp3 expression and resulting in the generation of pathogenic “ex-

Foxp3<sup>+</sup> cells which have been shown to induce diabetes within 8–11 days after transfer into NOD *Rag2*<sup>-/-</sup> mice [24;59]. This type of reversibility, or plasticity, of T cells is one of the major concerns with current T cell immunotherapies, in that beneficial effects may only be transient if cytokines, such as IL-6, act as a local barrier to the establishment of durable regulatory profiles.

In addition to this role as an inhibitor of Tregs, recent studies with human T cells have suggested that IL-6 may drive an important phenotype characteristic of treatment-refractory autoimmunity: In these studies, Teff from T1D subjects were found to be resistant to suppression by Treg from healthy controls [44]. Follow-up work demonstrated that this phenotype was also present in patients with some other autoimmune diseases and that this finding correlated with hyperactivity of the IL-6 signaling pathway, suggesting that autoreactive Teff are driven by tonic IL-6 signals to be refractory to regulation (Buckner et al., in press). So although IL-6 is often regarded as a lineage determinant for T cell development, it is more than that, with functional consequences promoting the establishment and maintenance of a proinflammatory, anti-regulatory tissue environment in autoimmunity.

As noted previously, the magnitude and activity of the Th17 pathway in T1D is a focus of active investigation. It has been suggested that the increase in Th17 cells observed in patients may be due to increased production of IL-6 by T1D monocytes [6], and this could also relate to increased Th17 activity upon T cell activation [20;31] and in the pancreas of T1D subjects [16].

Tocilizumab is a recombinant humanized monoclonal antibody specific for the human IL-6 receptor (IL-6R) that binds both the membrane-bound and soluble forms of IL-6R, currently approved for use in RA and sJIA in the USA and Europe. The pediatric experience is particularly promising for practical consideration of use in T1D, since tocilizumab has been used in young children with systemic juvenile idiopathic arthritis, yielding dramatic success, with approximately 90% of children achieving an ACR50 response. Tocilizumab has also shown preliminary efficacy in pilot open label phase 1 trials or small case studies in a variety of autoimmune or inflammatory conditions, including SLE, Takayasu and giant cell arteritis, Crohn's disease, systemic sclerosis, polymyositis, relapsing polychondritis, polymyalgia rheumatica, ankylosing spondylitis, and Behcet's disease [50]. In patients with RA after 3 infusions of tocilizumab, there was a decrease in the frequency of peripheral Th1 and Th17 cells and a corresponding increase of Treg [42], correlated with significant clinical response.

There have been no clinical studies in diabetes with tocilizumab. In a small open-label study of non-diabetic RA patients, tocilizumab therapy resulted in a significant decrease in the HOMA index for insulin resistance and a significant increase in serum adiponectin levels [45]. Also of note are the results of a recent mendelian randomization analysis for a SNP in the *IL6R* gene (Asp358Ala) that phenotypically recapitulates the effects of tocilizumab, which showed reduced risks for coronary heart disease and type 2 diabetes [19]. Thus, anti-IL-6R therapy may have metabolically favorable effects relating to the metabolic properties of subjects with T1D, notably insulin resistance. On the other hand, a recent study using sorted human islet cells suggested that IL-6 may have a role in stimulating alpha cell release of GLP-1 and thereby improve beta cell function [13].

### 3.0 Towards combination therapy including antigen delivery

Cytokine inhibition, as described in this article, is designed to play a major role in establishing a tissue milieu and cellular microenvironment permissive for regulatory immune responses, by simultaneously interfering with proinflammatory pathways and promoting alternative cellular functions that are more homeostatic. These cytokine

pathways, however, are intrinsically malleable, designed and honed by evolution to be flexible adaptors to external and tissue stress. As a consequence, one way to view cytokine inhibition in T1D is to view therapy as a staged process, in which a regulatory platform needs to be nurtured in vivo, suitable for enabling additional immune modulation therapy to succeed. Cytokine inhibition is an important approach for achieving this platform, but since the malleable nature of cytokine pathways requires them to be reversible and adaptable, it is unlikely that cytokine inhibition alone will be able to achieve durable therapeutic success.

Adaptive immunity to antigens, on the other hand, is designed to engender durable effector memory populations that can sustain an established immunological program. In this regard, a key goal for using cytokine inhibition as a therapeutic platform is to establish an in vivo milieu suitable for directing the adaptive immune response towards a therapeutically desirable outcome. For T1D, given current knowledge about effector pathways, blockade of cytokines such as IL-1, TNF, IL-12/23, and IL-6 has the potential to help create a suitable platform that will favor regulatory adaptive responses at the expense of proinflammatory Th1 and Th17 ones. A recent example is the ability of IL-1 blockade, in itself ineffective, to synergize with suboptimal doses of anti-CD3 mAb to accelerate and potentiate the reversal of overt T1D in NOD mice. [1].

An important question, however, is whether the autoantigen exposure that naturally occurs in T1D is sufficient to push the adaptive pathways towards such a regulatory and anti-inflammatory outcome. It is quite possible that it will be necessary to provide additional antigen, a form of therapeutic vaccination, simultaneously with cytokine blockade to assure the desired adaptive immune response. The T1D field has experimented with antigen therapies using various forms of insulin and recently an alum-formulated GAD vaccine, but these trials all lacked the fundamental pro-regulatory and anti-inflammatory platform discussed here. Indeed, some of these trials used adjuvants that may have actually interfered with prospective beneficial effects, and exploratory studies on tolerogenic forms of adjuvants are badly needed. Given the success of some of the anti-cytokine therapeutics in other autoimmune diseases, and the mechanistic rationale for simultaneously blocking proinflammatory effector pathways and promoting regulation, the time is at hand to move forward with additional trials using available cytokine inhibitors in T1D and pave the way for combining these treatments with antigen delivery.

## Acknowledgments

We thank the many members of the T1D immunotherapy community, who have contributed to the concepts discussed in this article, the JDRF and NIH for their support of T1D clinical trials, and colleagues at our institutions and elsewhere who have been instrumental in performing the studies cited herein.

## Abbreviations

<b>DAMPs</b>	damage/danger-associated molecular patterns
<b>ROS</b>	reactive oxygen species
<b>T1D</b>	type 1 diabetes
<b>T2D</b>	type 2 diabetes
<b>Treg</b>	regulatory T cell(s)
<b>Teff</b>	effector T cell(s)
<b>TLR</b>	toll-like receptor
<b>IL-1RA</b>	IL-1 receptor antagonist

<b>IL-1RAcP</b>	IL-1 receptor accessory protein
<b>TXNIP</b>	thioredoxin inhibitory protein
<b>LPS</b>	lipopolysaccharide
<b>NOD</b>	non-obese diabetic (mice)

## Reference List

1. Ablamunits V, Henegariu O, Hansen JB, Opare-Addo L, Preston-Hurlburt P, Santamaria P, Mandrup-Poulsen T, Herold KC. Synergistic reversal of type 1 diabetes in NOD mice with anti-CD3 and interleukin-1 blockade: evidence of improved immune regulation. *Diabetes*. 2012; 61:145–154. [PubMed: 22043003]
2. Arif S, Moore F, Marks K, Bouckennooghe T, Dayan CM, Planas R, Vives-Pi M, Powrie J, Tree T, Marchetti P, Huang GC, Gurzov EN, Pujol-Borrell R, Eizirik DL, Peakman M. Peripheral and islet interleukin-17 pathway activation characterizes human autoimmune diabetes and promotes cytokine-mediated beta-cell death. *Diabetes*. 2011; 60:2112–2119. [PubMed: 21659501]
3. Bending D, De la Pena H, Veldhoen M, Phillips JM, Uyttenhove C, Stockinger B, Cooke A. Highly purified Th17 cells from BDC2.5NOD mice convert into Th1-like cells in NOD/SCID recipient mice. *J Clin Invest*. 2009; 119:565–572. [PubMed: 19188681]
4. Benson JM, Peritt D, Scallan BJ, Heavner GA, Shealy DJ, Giles-Komar JM, Mascelli MA. Discovery and mechanism of ustekinumab: a human monoclonal antibody targeting interleukin-12 and interleukin-23 for treatment of immune-mediated disorders. *MABs*. 2011; 3:535–545. [PubMed: 22123062]
5. Bettelli E, Carrier Y, Gao W, Korn T, Strom TB, Oukka M, Weiner HL, Kuchroo VK. Reciprocal developmental pathways for the generation of pathogenic effector TH17 and regulatory T cells. *Nature*. 2006; 441:235–238. [PubMed: 16648838]
6. Bradshaw EM, Raddassi K, Elyaman W, Orban T, Gottlieb PA, Kent SC, Hafler DA. Monocytes from patients with type 1 diabetes spontaneously secrete proinflammatory cytokines inducing Th17 cells. *J Immunol*. 2009; 183:4432–4439. [PubMed: 19748982]
7. Cavelti-Weder C, Babians-Brunner A, Keller C, Stahel MA, Kurz-Levin M, Zayed H, Solinger AM, Mandrup-Poulsen T, Dinarello CA, Donath MY. Effects of gevokizumab on glycemia and inflammatory markers in type 2 diabetes. *Diabetes Care*. 2012; 35:1654–1662. [PubMed: 22699287]
8. Croxtall JD. Ustekinumab: a review of its use in the management of moderate to severe plaque psoriasis. *Drugs*. 2011; 71:1733–1753. [PubMed: 21902296]
9. Dinarello CA. The many worlds of reducing interleukin-1. *Arthritis Rheum*. 2005; 52:1960–1967. [PubMed: 15986340]
10. Dinarello CA. Immunological and inflammatory functions of the interleukin-1 family. *Annu Rev Immunol*. 2009; 27:519–550. [PubMed: 19302047]
11. Dinarello CA. Interleukin-1 in the pathogenesis and treatment of inflammatory diseases. *Blood*. 2011; 117:3720–3732. [PubMed: 21304099]
12. Ehses JA, Lacraz G, Giroix MH, Schmidlin F, Coulaud J, Kassis N, Irminger JC, Kergoat M, Portha B, Homo-Delarche F, Donath MY. IL-1 antagonism reduces hyperglycemia and tissue inflammation in the type 2 diabetic GK rat. *Proc Natl Acad Sci USA*. 2009; 106:13998–14003. [PubMed: 19666548]
13. Ellingsgaard H, Hauselmann I, Schuler B, Habib AM, Baggio LL, Meier DT, Eppler E, Bouzakri K, Wueest S, Muller YD, Hansen AM, Reinecke M, Konrad D, Gassmann M, Reimann F, Halban PA, Gromada J, Drucker DJ, Gribble FM, Ehses JA, Donath MY. Interleukin-6 enhances insulin secretion by increasing glucagon-like peptide-1 secretion from L cells and alpha cells. *Nat Med*. 2011; 17:1481–1489. [PubMed: 22037645]
14. Elliott M, Benson J, Blank M, Brodmerkel C, Baker D, Sharples KR, Szapary P. Ustekinumab: lessons learned from targeting interleukin-12/23p40 in immune-mediated diseases. *Ann NY Acad Sci*. 2009; 1182:97–110. [PubMed: 20074279]

15. Emamaullee JA, Davis J, Merani S, Toso C, Elliott JF, Thiesen A, Shapiro AM. Inhibition of Th17 cells regulates autoimmune diabetes in NOD mice. *Diabetes*. 2009; 58:1302–1311. [PubMed: 19289457]
16. Ferraro A, Socci C, Stabilini A, Valle A, Monti P, Piemonti L, Nano R, Olek S, Maffi P, Scavini M, Secchi A, Staudacher C, Bonifacio E, Battaglia M. Expansion of Th17 cells and functional defects in T regulatory cells are key features of the pancreatic lymph nodes in patients with type 1 diabetes. *Diabetes*. 2011; 60:2903–2913. [PubMed: 21896932]
17. Fujihira K, Nagata M, Moriyama H, Yasuda H, Arisawa K, Nakayama M, Maeda S, Kasuga M, Okumura K, Yagita H, Yokono K. Suppression and acceleration of autoimmune diabetes by neutralization of endogenous interleukin-12 in NOD mice. *Diabetes*. 2000; 49:1998–2006. [PubMed: 11118000]
18. Herold KC, Gitelman SE, Masharani U, Hagopian W, Bisikirska B, Donaldson D, Rother K, Diamond B, Harlan DM, Bluestone JA. A single course of anti-CD3 monoclonal antibody hOKT3gamma1(Ala-Ala) results in improvement in C-peptide responses and clinical parameters for at least 2 years after onset of type 1 diabetes. *Diabetes*. 2005; 54:1763–1769. [PubMed: 15919798]
19. Hingorani AD, Casas JP. The interleukin-6 receptor as a target for prevention of coronary heart disease: a mendelian randomisation analysis. *Lancet*. 2012; 379:1214–1224. [PubMed: 22421340]
20. Honkanen J, Nieminen JK, Gao R, Luopajarvi K, Salo HM, Ilonen J, Knip M, Otonkoski T, Vaarala O. IL-17 immunity in human type 1 diabetes. *J Immunol*. 2010; 185:1959–1967. [PubMed: 20592279]
21. Joseph J, Bittner S, Kaiser FM, Wiendl H, Kissler S. IL-17 silencing does not protect nonobese diabetic mice from autoimmune diabetes. *J Immunol*. 2012; 188:216–221. [PubMed: 22116823]
22. Keymeulen B, Walter M, Mathieu C, Kaufman L, Gorus F, Hilbrands R, Vandemeulebroucke E, Van d, Crenier VL, De BC, Candon S, Waldmann H, Ziegler AG, Chatenoud L, Pipeleers D. Four-year metabolic outcome of a randomised controlled CD3-antibody trial in recent-onset type 1 diabetic patients depends on their age and baseline residual beta cell mass. *Diabetologia*. 2010; 53:614–623. [PubMed: 20225393]
23. Korn T, Mitsdoerffer M, Croxford AL, Awasthi A, Dardalhon VA, Galileos G, Vollmar P, Stritesky GL, Kaplan MH, Waisman A, Kuchroo VK, Oukka M. IL-6 controls Th17 immunity in vivo by inhibiting the conversion of conventional T cells into Foxp3+ regulatory T cells. *Proc Natl Acad Sci USA*. 2008; 105:18460–18465. [PubMed: 19015529]
24. Lal G, Zhang N, van der Touw W, Ding Y, Ju W, Bottinger EP, Reid SP, Levy DE, Bromberg JS. Epigenetic regulation of Foxp3 expression in regulatory T cells by DNA methylation. *J Immunol*. 2009; 182:259–273. [PubMed: 19109157]
25. Larsen CM, Faulenbach M, Vaag A, Volund A, Eshes JA, Seifert B, Mandrup-Poulsen T, Donath MY. Interleukin-1-receptor antagonist in type 2 diabetes mellitus. *N Engl J Med*. 2007; 356:1517–1526. [PubMed: 17429083]
26. Leonardi CL, Kimball AB, Papp KA, Yeilding N, Guzzo C, Wang Y, Li S, Dooley LT, Gordon KB. Efficacy and safety of ustekinumab, a human interleukin-12/23 monoclonal antibody, in patients with psoriasis: 76-week results from a randomised, double-blind, placebo-controlled trial (PHOENIX 1). *Lancet*. 2008; 371:1665–1674. [PubMed: 18486739]
27. Mandrup-Poulsen T. The role of interleukin-1 in the pathogenesis of IDDM. *Diabetologia*. 1996; 39:1005–1029. [PubMed: 8877284]
28. Mandrup-Poulsen T, Bendtzen K, Dinarello CA, Nerup J. Human tumor necrosis factor potentiates human interleukin 1-mediated rat pancreatic beta-cell cytotoxicity. *J Immunol*. 1987; 139:4077–4082. [PubMed: 3320203]
29. Mandrup-Poulsen T, Pickersgill L, Donath MY. Blockade of interleukin 1 in type 1 diabetes mellitus. *Nat Rev Endocrinol*. 2010; 6:158–166. [PubMed: 20173777]
30. Martinon F, Mayor A, Tschopp J. The inflammasomes: guardians of the body. *Annu Rev Immunol*. 2009; 27:229–265. [PubMed: 19302040]
31. Marwaha AK, Crome SQ, Panagiotopoulos C, Berg KB, Qin H, Ouyang Q, Xu L, Priatel JJ, Levings MK, Tan R. Cutting edge: Increased IL-17-secreting T cells in children with new-onset type 1 diabetes. *J Immunol*. 2010; 185:3814–3818. [PubMed: 20810982]

32. Mastrandrea L, Yu J, Behrens T, Buchlis J, Albini C, Fournier S, Quattrin T. Etanercept treatment in children with new-onset type 1 diabetes: pilot randomized, placebo-controlled, double-blind study. *Diabetes Care*. 2009; 32:1244–1249. [PubMed: 19366957]
33. Murakami M, Nishimoto N. The value of blocking IL-6 outside of rheumatoid arthritis: current perspective. *Curr Opin Rheumatol*. 2011; 23:273–277. [PubMed: 21427577]
34. Nicoletti F, Di MR, Zaccone P, Magro G, Di MM, Grasso S, Meroni PL. Endogenous interleukin-12 only plays a key pathogenetic role in non-obese diabetic mouse diabetes during the very early stages of the disease. *Immunology*. 1999; 97:367–370. [PubMed: 10447755]
35. Orban T, Bundy B, Becker DJ, DiMeglio LA, Gitelman SE, Goland R, Gottlieb PA, Greenbaum CJ, Marks JB, Monzavi R, Moran A, Raskin P, Rodriguez H, Russell WE, Schatz D, Wherrett D, Wilson DM, Krischer JP, Skyler JS. Co-stimulation modulation with abatacept in patients with recent-onset type 1 diabetes: a randomised, double-blind, placebo-controlled trial. *Lancet*. 2011; 378:412–419. [PubMed: 21719096]
36. Osłowski CM, Hara T, O’Sullivan-Murphy B, Kanekura K, Lu S, Hara M, Ishigaki S, Zhu LJ, Hayashi E, Hui ST, Greiner D, Kaufman RJ, Bortell R, Urano F. Thioredoxin-interacting protein mediates ER stress-induced beta cell death through initiation of the inflammasome. *Cell Metab*. 2012; 16:265–273. [PubMed: 22883234]
37. Papp KA, Langley RG, Lebwohl M, Krueger GG, Szapary P, Yeilding N, Guzzo C, Hsu MC, Wang Y, Li S, Dooley LT, Reich K. Efficacy and safety of ustekinumab, a human interleukin-12/23 monoclonal antibody, in patients with psoriasis: 52-week results from a randomised, double-blind, placebo-controlled trial (PHOENIX 2). *Lancet*. 2008; 371:1675–1684. [PubMed: 18486740]
38. Pescovitz MD, Greenbaum CJ, Krause-Steinrauf H, Becker DJ, Gitelman SE, Goland R, Gottlieb PA, Marks JB, McGee PF, Moran AM, Raskin P, Rodriguez H, Schatz DA, Wherrett D, Wilson DM, Lachin JM, Skyler JS. Rituximab, B-lymphocyte depletion, and preservation of beta-cell function. *N Engl J Med*. 2009; 361:2143–2152. [PubMed: 19940299]
39. Rose-John S, Waetzig GH, Scheller J, Grotzinger J, Seegert D. The IL-6/sIL-6R complex as a novel target for therapeutic approaches. *Expert Opin Ther Targets*. 2007; 11:613–624. [PubMed: 17465721]
40. Rothe H, O’Hara RM Jr, Martin S, Kolb H. Suppression of cyclophosphamide induced diabetes development and pancreatic Th1 reactivity in NOD mice treated with the interleukin (IL)-12 antagonist IL-12(p40)2. *Diabetologia*. 1997; 40:641–646. [PubMed: 9222642]
41. Sakata M, Yasuda H, Moriyama H, Yamada K, Kotani R, Kurohara M, Okumachi Y, Kishi M, Arai T, Hara K, Hamada H, Yokono K, Nagata M. Prevention of recurrent but not spontaneous autoimmune diabetes by transplanted NOD islets adenovirally transduced with immunomodulating molecules. *Diabetes Res Clin Pract*. 2008; 80:352–359. [PubMed: 18400329]
42. Samson M, Audia S, Janikashvili N, Ciudad M, Trad M, Fraszczak J, Ornetti P, Maillefert JF, Miossec P, Bonnotte B. Brief report: inhibition of interleukin-6 function corrects Th17/Treg cell imbalance in patients with rheumatoid arthritis. *Arthritis Rheum*. 2012; 64:2499–2503. [PubMed: 22488116]
43. Sandborn WJ, Gasink C, Gao LL, Blank MA, Johanns J, Guzzo C, Sands BE, Hanauer SB, Targan S, Rutgeerts P, Ghosh S, de Villiers WJ, Panaccione R, Greenberg G, Schreiber S, Lichtiger S, Feagan BG. Ustekinumab induction and maintenance therapy in refractory Crohn’s disease. *N Engl J Med*. 2012; 367:1519–1528. [PubMed: 23075178]
44. Schneider A, Rieck M, Sanda S, Pihoker C, Greenbaum C, Buckner JH. The effector T cells of diabetic subjects are resistant to regulation via CD4+ FOXP3+ regulatory T cells. *J Immunol*. 2008; 181:7350–7355. [PubMed: 18981158]
45. Schultz O, Oberhauser F, Saech J, Rubbert-Roth A, Hahn M, Krone W, Laudes M. Effects of inhibition of interleukin-6 signalling on insulin sensitivity and lipoprotein (a) levels in human subjects with rheumatoid diseases. *PLoS ONE*. 2010; 5:e14328. [PubMed: 21179199]
46. Suarez-Farinas M, Li K, Fuentes-Duculan J, Hayden K, Brodmerkel C, Krueger JG. Expanding the psoriasis disease profile: interrogation of the skin and serum of patients with moderate-to-severe psoriasis. *J Invest Dermatol*. 2012; 132:2552–2564. [PubMed: 22763790]

47. Sumpter KM, Adhikari S, Grishman EK, White PC. Preliminary studies related to anti-interleukin-1beta therapy in children with newly diagnosed type 1 diabetes. *Pediatr Diabetes*. 2011; 12:656–667. [PubMed: 21518168]
48. Sun M, Fink PJ. A new class of reverse signaling costimulators belongs to the TNF family. *J Immunol*. 2007; 179:4307–4312. [PubMed: 17878324]
49. Sutherland AP, Van BT, Wurster AL, Suto A, Michaud M, Zhang D, Grusby MJ, von HM. Interleukin-21 is required for the development of type 1 diabetes in NOD mice. *Diabetes*. 2009; 58:1144–1155. [PubMed: 19208913]
50. Tanaka T, Narazaki M, Kishimoto T. Therapeutic targeting of the interleukin-6 receptor. *Annu Rev Pharmacol Toxicol*. 2012; 52:199–219. [PubMed: 21910626]
51. Trembleau S, Penna G, Bosi E, Mortara A, Gately MK, Adorini L. Interleukin 12 administration induces T helper type 1 cells and accelerates autoimmune diabetes in NOD mice. *J Exp Med*. 1995; 181:817–821. [PubMed: 7836934]
52. Trembleau S, Penna G, Gregori S, Gately MK, Adorini L. Deviation of pancreas-infiltrating cells to Th2 by interleukin-12 antagonist administration inhibits autoimmune diabetes. *Eur J Immunol*. 1997; 27:2330–2339. [PubMed: 9341777]
53. Trembleau S, Penna G, Gregori S, Giarratana N, Adorini L. IL-12 administration accelerates autoimmune diabetes in both wild-type and IFN-gamma-deficient nonobese diabetic mice, revealing pathogenic and protective effects of IL-12-induced IFN-gamma. *J Immunol*. 2003; 170:5491–5501. [PubMed: 12759426]
54. Vendrame F, Pileggi A, Laughlin E, Allende G, Martin-Pagola A, Molano RD, Diamantopoulos S, Standifer N, Geubtner K, Falk BA, Ichii H, Takahashi H, Snowwhite I, Chen Z, Mendez A, Chen L, Sageshima J, Ruiz P, Ciancio G, Ricordi C, Reijonen H, Nepom GT, Burke GW III, Pugliese A. Recurrence of type 1 diabetes after simultaneous pancreas-kidney transplantation, despite immunosuppression, is associated with autoantibodies and pathogenic autoreactive CD4 T-cells. *Diabetes*. 2010; 59:947–957. [PubMed: 20086230]
55. Yasuda H, Nagata M, Arisawa K, Yoshida R, Fujihira K, Okamoto N, Moriyama H, Miki M, Saito I, Hamada H, Yokono K, Kasuga M. Local expression of immunoregulatory IL-12p40 gene prolonged syngeneic islet graft survival in diabetic NOD mice. *J Clin Invest*. 1998; 102:1807–1814. [PubMed: 9819366]
56. Zaghi D, Krueger GG, Callis DK. Ustekinumab: a review in the treatment of plaque psoriasis and psoriatic arthritis. *J Drugs Dermatol*. 2012; 11:160–167. [PubMed: 22270196]
57. Zhang J, Huang Z, Sun R, Tian Z, Wei H. IFN-gamma induced by IL-12 administration prevents diabetes by inhibiting pathogenic IL-17 production in NOD mice. *J Autoimmunity*. 2012; 38:20–28. [PubMed: 22186068]
58. Zhou R, Tardivel A, Thorens B, Choi I, Tschopp J. Thioredoxin-interacting protein links oxidative stress to inflammasome activation. *Nat Immunol*. 2010; 11:136–140. [PubMed: 20023662]
59. Zhou X, Bailey-Bucktrout SL, Jeker LT, Penaranda C, Martinez-Llordella M, Ashby M, Nakayama M, Rosenthal W, Bluestone JA. Instability of the transcription factor Foxp3 leads to the generation of pathogenic memory T cells in vivo. *Nat Immunol*. 2009; 10:1000–1007. [PubMed: 19633673]

### Highlights

- cytokine inhibition supports regulatory immunological mechanisms
- cytokines influence immunologic determinism in tissue microenvironments
- cytokine blockade can establish a foundation for antigen-specific therapy
- several cytokine inhibitors should be evaluated in T1D clinical trials

**Table 1**

## The interleukin-1 family

Nomenclature		Receptor	Receptor accessory protein	Decoy receptor
IL-1F1	IL-1 <sup>+</sup>	IL-1RI	IL-1RAcP	IL-1RII
IL-1F2	IL-1 <sup>*</sup>	IL-1RI	IL-1RAcP	IL-1RII
IL-1F3	<i>IL-1Ra</i>	IL-1RI		IL-1RII
IL-1F6	IL-36	IL-1Rrp2	IL-1RAcP	
IL-1F8	IL-36	IL-1Rrp2	IL-1RAcP	
IL-1F9	IL-36	IL-1Rrp2	IL-1RAcP	
IL-1F5	<i>IL-36Ra</i>	IL-1Rrp2		
IL-1F4	IL-18 <sup>*</sup>	IL-18R	IL-18R	
IL-1F7	<i>IL-37a,b<sup>+</sup></i>	IL-18R	?	
IL-1F11	IL-33 <sup>*+</sup>	ST2	IL-1RAcP	
IL-1F10	<i>IL-38</i>	?	?	

*italics* indicate antagonists or partial agonists

<sup>+</sup> also transcriptional co-factors

<sup>\*</sup> generated from inflammasome processing by caspase-1

**Table 2**

## The tumor necrosis factor family

Nomenclature	Receptor	Decoy receptor
membrane/soluble TNF	p55, p75 TNF-R, SFV T2	
Lymphotoxin / 1, 2	p55, p75 TNF-R, LT R	
LIGHT	LT R, DCR2, HVEM	
CD40L	CD40	
FasL	Fas	
TRAIL	TRAIL R1-2	TRAIL 3-4
APRIL	BCMA, TAC1	
BAFF/Blyss	BAFF-R/BL-3, BCMA, TAC1	
CD27L	CD27	
CD30L	CD30	
OX40L	OX40	
4-1 BBL	4-1 B-R	
GITRL	GITR	
RANKL	RANK, OPG	
TWEAK	TWEAK-R	
EDA	EDA-R	